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SYNTHESIS OF PEPTIDE NUCLEIC ACID MONOMERS

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ABSTRACT: The chemical synthesis of peptide nucleic acid (PNA) monomers is described using Fmoc (backbone), anisoyl (cytosine, adenine), 4-*tert*-butylbenzoyl (cytosine) and isobutyryl/diphenylcarbamoyl (guanine) protecting group combinations. For the guanine monomer the alkylation was realized both in a Mitsunobu [DIAD, triphenylphosphine or (4-dimethylaminophenyl)diphenylphosphine, *tert*-butyl glycolate] and in a low-temperature, sodium-hydride mediated alkylation (*tert*-butyl bromoacetate) to give the *N*⁹-substituted derivative.

Peptide nucleic acids (PNA) are oligonucleotide analogues in which the sugar-phosphodiester backbone is replaced by a nucleobase-derivatized *N*-(2-aminoethyl)glycine chain. PNA oligomers have a number of properties (*e.g.* DNA and RNA recognizing ability) which make them very useful in antisense therapeutics and as diagnostics tools.

Until recently the assembly of PNA oligomers was based on a Merrifield solid-phase synthesis of Boc/Z-protected monomers¹. Relatively few experience is available with other combination of protecting groups (*e.g.* Fmoc/Z², monomethoxytrityl/anisoyl³) and there is a continuous need for deprotection conditions compatible with different protocols.

We would like to report herein on our results concerning the synthesis of PNA monomers (1-5, FIG) using Fmoc group for the backbone and various other groups for the heterobase *N*-protection in order to enable the oligomer synthesis to be performed on a peptide synthesizer using Fmoc strategy. Nucleobase-substituted acetic acids were

prepared as follows. Thymin-1-ylacetic acid was prepared according to a published procedure¹. Cytosine was anisoylated and subsequently alkylated to give the corresponding acid. As this compound was very poorly soluble in most organic solvents the more lipophilic 4-*tert*-butylbenzoylated derivative was alkylated and the acid was obtained using the procedure by Will *et al.*³. The anisoylated adenine derivative was similarly prepared³. However, we have found more convenient to peranisoylate and then hydrolyze *N*⁹-ethoxycarbonyladenine to afford the same acid.

For the guanine monomer *N*²-isobutyryl-*O*⁶-diphenylcarbamoylguanine was chosen following the methodology by Zou and Robins^{4,5} in order to avoid the formation of regioisomers. The persilylated derivative of the above compound can effectively be glycosylated or alkylated with alkoxymethyl halides⁵ at *N*⁹ but it does not undergo reaction with *tert*-butyl bromoacetate neither in the absence nor in the presence of mercury(II) cyanide^{6,7}. We then investigated the Mitsunobu reaction⁸ of *N*²-isobutyryl-*O*⁶-diphenylcarbamoylguanine with *tert*-butyl glycolate⁹. The resulting ester was obtained along with a negligible amount of the *N*⁷ regioisomer. It was difficult to remove completely the triphenylphosphine oxide byproduct therefore (4-dimethylaminophenyl)-diphenylphosphine¹⁰ was used which gave an acid-removable phosphine oxide¹¹. Alternatively, low-temperature, sodium hydride-mediated alkylation with *tert*-butyl bromoacetate gave the product with excellent *N*⁹/*N*⁷ selectivity. When the alkylation was performed near room temperature significant amounts of *N*⁷ isomer was formed. The free acid was then obtained in an acidic hydrolysis of the *tert*-butyl group.

The coupling of the appropriate nucleobase-substituted acetic acids with the common backbone *tert*-butyl *N*-[2-(*N*-9-fluorenylmethoxycarbonyl)aminoethyl]glycinate hydrochloride² was performed using standard methods of peptide synthesis.

The synthesis of some new PNA oligomers from our monomers is under way.

REFERENCES

1. Dueholm, K. L.; Egholm, M.; Behrens, C.; Christensen, L.; Hansen, H. F.; Vulpius, T.; Petersen, K. H.; Berg, R. H.; Nielsen, P. E.; Buchardt, O., *J. Org. Chem.*, **1994**, *59*, 5767-5773.
2. Thomson, S. A.; Josey, J. A.; Cadilla, R.; Gaul, M. D.; Hassman, C. F.; Luzzio, M. J.; Pipe, A. J.; Reed, K. L.; Ricca, D. J.; Wiethe, R. W.; Noble, S. A., *Tetrahedron*, **1995**, *51*, 6179-6194.

3. Will, D. W.; Langner, D.; Knolle, J.; Uhlmann, E., *Tetrahedron*, **1995**, *51*, 12069-12082.
4. Zou, R.; Robins, M. J., *Can. J. Chem.*, **1987**, *65*, 1436-1437.
5. Robins, M. J.; Zou, R. M.; Guo, Z. Q.; Wnuk, S. F., *J. Org. Chem.*, **1996**, *61*, 9207-9212.
6. Kim, C. U.; Misco, P. F.; Luh, B. Y.; Martin, J. C., *Tetrahedron Lett.*, **1990**, *31*, 3257-3260.
7. Kim, C. U.; Misco, P. F.; Luh, B. Y.; Hitchcock, M. J. M.; Ghazzouli, I.; Martin, J. C., *J. Med. Chem.*, **1991**, *34*, 2286-2294.
8. Jenny, T. F.; Schneider, K. C.; Benner, S. A., *Nucleosides Nucleotides*, **1992**, *11*, 1257-1261.
9. Kricheldorf, H. R.; Kaschig, J., *Liebigs Ann. Chem.*, **1976**, 882-890.
10. Brune, H. A.; Falck, M.; Hemmer, R.; Schmidtberg, G.; Alt, H. G., *Chem. Ber.*, **1984**, *117*, 2791-2802.
11. von Itzstein, M.; Mocerino, M., *Synth. Commun.*, **1990**, *20*, 2049-2057.